

Influences of free-air CO₂ enrichment (FACE), nitrogen fertilizer and crop residue incorporation on CH₄ emissions from irrigated rice fields

Baohua Xie · Zaixing Zhou · Baoling Mei · Xunhua Zheng ·
Haibo Dong · Rui Wang · Shenghui Han · Feng Cui ·
Yinghong Wang · Jianguo Zhu

Received: 22 November 2011 / Accepted: 27 July 2012 / Published online: 5 August 2012
© Springer Science+Business Media B.V. 2012

Abstract To investigate the response of methane (CH₄) emissions to an elevated atmospheric carbon dioxide (CO₂) concentration (200 ± 40 μmol mol⁻¹ higher than the ambient atmosphere), we performed a 4-year multi-factorial experiment at a subtropical rice paddy that contained sandy loam soil in the Yangtze River Delta from 2004 to 2007 using free-air CO₂ enrichment (FACE) technology. Our results revealed that the elevated atmospheric CO₂ increased the seasonal cumulative CH₄ emissions by 15 % on average during the 4-year period. The increase was insignificant and much weaker than the previous

studies, which might be primarily attributed to the absence of a significant difference in the rice biomass between the two CO₂ levels in half of the field treatments. Crop residue incorporation hindered the stimulatory effects induced by the elevated CO₂, which were 37, 14 and 6 % for the fields that were incorporated with none, half or all of the wheat straws that were harvested in the preceding winter wheat season, respectively. Nitrogen fertilizers application also hindered the stimulatory effects of the elevated CO₂ on the CH₄ emissions. The CO₂ stimulatory effect was 39 % for the field without nitrogen fertilizers, and reduced to 17, 7 and 5 % for the field with nitrogen fertilization of 125, 250 and 350 kg N ha⁻¹, respectively. The regulation of nitrogen fertilizers on the CO₂ effects in this experiment does not well agree with the previous studies, which might be because the soil type was different from those of the previous studies. Thus, further studies are necessary to evaluate the role of soil properties in regulating the effects of elevated atmospheric CO₂ on CH₄ emissions from managed and natural wetlands. There were no significant interactions between the atmospheric CO₂ and the incorporations of nitrogen fertilizer and crop residue. Appropriate experiments are necessary for better understanding of the interact influences of the elevated CO₂ and farm managements.

B. Xie · Z. Zhou · B. Mei · X. Zheng (✉) ·
H. Dong · R. Wang · S. Han · F. Cui · Y. Wang
State Key Laboratory of Atmospheric Boundary Layer
Physics and Atmospheric Chemistry, Institute of
Atmospheric Physics, Chinese Academy of Sciences,
Beijing 100029, China
e-mail: xunhua.zheng@post.iap.ac.cn

B. Xie
Laboratory of Coastal Wetland Ecology, Key Laboratory
of Coastal Zone Environmental Process, Shandong
Provincial Key Laboratory of Coastal Zone
Environmental Process, Yantai Institute of Coastal Zone
Research, Chinese Academy of Sciences (CAS), Yantai
264003, People's Republic of China

J. Zhu
State Key Laboratory of Soil and Sustainable Agriculture,
Institute of Soil Sciences, Chinese Academy of Sciences,
Nanjing 210008, China

Keywords Carbon dioxide · Methane · Emission ·
Free-air CO₂ enrichment (FACE) · Rice field ·
Nitrogen · Crop residue

Introduction

Despite the dedicated efforts of scientists and policy-makers in many countries, anthropogenic greenhouse gas emissions still increased at a marked rate over the last two decades (IPCC 2007). Methane (CH₄), which is the second most important greenhouse gas, contributes to approximately 20 % of the anthropogenic global warming effect (Houghton et al. 2001). Paddy rice fields account for approximately 5–19 % of the total global CH₄ emissions to the atmosphere (IPCC 2007). The intensification of rice production is imperative in meeting the increasing demand for rice by the increasing human population (Cassman et al. 1998). This demand will most likely lead to increased CH₄ emissions if no appropriate mitigation strategies are developed and applied. The increasing atmospheric carbon dioxide (CO₂) concentration is hypothesized to amplify the level of CH₄ that is released from not only natural (Dacey et al. 1994; Hutchin et al. 1995) but also managed wetlands, including irrigated rice fields (Inubushi et al. 2003; Tokida et al. 2010; Zheng et al. 2006; Ziska et al. 1998).

Empirical records provide incontestable evidence of global changes. The foremost among these changes is the increasing atmospheric CO₂ concentration, which has been predicted to double by the mid-twentyfirst century compared to the concentration in the pre-industrial era (IPCC 2001). Many studies have demonstrated a positive effect of elevated CO₂ levels on rice biomass (above and below ground) and grain yield (Kim et al. 2001, 2003; Yang et al. 2007). Elevated CO₂ levels also increase the soil microbial carbon levels, and they accelerate the turnover rate of the soil microbial carbon during the middle and later parts of the rice growing season (Cheng et al. 2001; Hoque et al. 2001). The stimulation of rice root biomass and tiller number by elevated CO₂ is expected to indirectly affect the CH₄ production and emission from paddy rice fields. Based on a few experiments that were performed under environmentally controlled conditions, such as in open top chambers (Dacey et al. 1994) or greenhouses (Yagi et al. 2002), an atmospheric CO₂ level that was elevated by 60–100 % above the ambient level significantly increased the CH₄ emission (by 19–300 %) in temperate (Cheng et al. 2006; Yagi et al. 2002) and tropical rice paddies (Allen et al. 2003; Ziska et al. 1998).

Free-air CO₂ enrichment (FACE) techniques have been developed in the past two decades (Kimball 1992; Okada et al. 2001) and have recently been applied to investigate the effects of the atmospheric CO₂ enrichment on the CH₄ emissions from managed (Inubushi et al. 2003; Tokida et al. 2010; Zheng et al. 2006) and natural (Silvola et al. 2003) wetland ecosystems. The FACE techniques can facilitate undisturbed field conditions and produce more reliable measurements in comparison with the use of environmentally controlled facilities (Long et al. 2006; Okada et al. 2001). A long-term FACE study of temperate paddy rice fields in Japan showed increases (26–51 %) in CH₄ emissions as a result of long-term CO₂ enrichment (Inubushi et al. 2003; Tokida et al. 2010). Another FACE study of subtropical paddy rice fields that were subjected to a rice–wheat rotation regime in China showed a significant stimulatory effect (53–188 %) on CH₄ emissions without or with a medium addition rate (1,000 kg C ha⁻¹) of wheat straw, but there was no significant effect when the conditions included a high addition rate (2,000 kg C ha⁻¹) of wheat straw (Zheng et al. 2006). The FACE studies that were simultaneously performed at five sites along a transect of oligotrophic peatland mires in Europe demonstrated no significant CO₂ effects on CH₄ emissions (Silvola et al. 2003). The discrepancy among these studies may be partly due to differences in experimental conditions, including different climates, soil types and farm management. In fact, CH₄ emissions from wetland ecosystems are directly controlled by a series of biochemical and microbial processes that are regulated by soil types and water regimes (Conrad 2002; Schimel 2000).

Nitrogenous fertilizers are commonly utilized in rice cultivation to achieve high grain yields. There is an ongoing discussion regarding the possible effects of nitrogen application on the CH₄ emission from rice fields. The application of nitrogen may directly or indirectly influence the processes that regulate CH₄ emission from paddy rice fields, which include CH₄ production, oxidation, and transport from the soil to the atmosphere. Nitrogen application not only stimulates the growth and activity of methane oxidizing bacteria (Bodelier and Laanbroek 2004; Schimel 2000), but also enhance the growth and activity of methanogenic bacteria (Cai et al. 2007). Nitrogen application would also affect CH₄ production and transport through the effect on rice growth because

rice plants can provide organic substrates and a major pathway transporting CH₄ from the paddy soil to the atmosphere (Huang et al. 2004; Kludze et al. 1993; Le Mer and Roger 2001). Because of the complexity and counter-balance among the different effects, it is still difficult to assess the net nitrogen effect on CH₄ emissions from rice fields.

Crop residue amendments have been commonly utilized to improve the soil fertility for rice production in China. The incorporation of crop residues provides a source of readily available carbon, which is believed to induce the higher release of CH₄ from rice paddies (Denier van der Gon and Neue 1995; Wang et al. 1999). The decomposition of incorporated organic material is the predominant source of methanogenic substrates in the early stage of the rice-growing season (Huang et al. 2004).

The incorporation of organic matter and nitrogen fertilizer may regulate the effect of an elevated CO₂ on CH₄ emissions from paddy rice fields. Zheng et al. (2006) reported that the impacts of the long-term atmospheric CO₂ enrichment on the CH₄ emissions from nitrogen-rich paddy rice ecosystems correlated positively with the nitrogen addition rates but negatively with the addition rates of decomposable organic carbon; both of the correlations were related to the soil nitrogen availability. However, studies that focus on the regulation of different farm management practices on the effects of elevated atmospheric CO₂ and the CH₄ emissions from rice fields are still lacking.

Currently, only three FACE studies of paddy rice ecosystem have been reported. All of the previous studies were conducted in clay paddies that utilized flooding water during the rice growing period, with the exception of one or two explicit mid-season drainages (Inubushi et al. 2003; Tokida et al. 2010; Xu et al. 2004; Zheng et al. 2006). Only one of the research projects performed a preliminary study of the simultaneous influences of elevated atmospheric CO₂, nitrogen fertilizer and organic matter amendments on CH₄ emissions. Furthermore, the response of CH₄ emissions from paddy rice ecosystems to the elevated CO₂ likely varies for different soil types. However, experimental evidence is still lacking. To enhance our understanding of the response of CH₄ emissions to elevated CO₂ conditions and different farm management practices, we performed a 4-year multi-factorial experiment using a FACE platform on a subtropical paddy rice field that was subjected to a rice–wheat

rotation. This experimental site was located on a sandy loam soil in the Yangtze River Delta. Here, we report the results of this experiment.

Materials and methods

Experimental site and rice cultivation

The experimental platform of free-air CO₂ enrichment (FACE) was established over a rice–winter wheat rotation ecosystem located in a suburb (32°35'N, 119°42'E) of Jiangdu county in Jiangsu province, which is located in the Yangtze River Delta region. The crop rotation pattern accounts for approximately 60 % of the regional paddy area (Zheng et al. 2000), and it is typical for other regions that have a similar climate, such as in eastern, central and southwestern China (Zheng et al. 2000; Zheng et al. 2006). This region displays a north subtropical monsoon climate. The annual precipitation is 918–978 mm, and 60 % of the precipitation occurs from June to September. The annual mean air temperature is 14.9 °C, and the annual frost-free period is approximately 220 days. The soil at the experimental site is Shajiang-Aquic Cambosols (CRGCST-Cooperative Research Group on Chinese Soil Taxonomy, 2001) with a sandy loam texture. The relevant soil properties at a 0–15 cm depth are as follows: tillage layer depth 13.0 cm, bulk density 1.16 g cm⁻³, soil organic carbon 18.4 g kg⁻¹, total nitrogen 1.45 g kg⁻¹, total phosphorous 0.63 g kg⁻¹, total potassium 14.0 g kg⁻¹, available phosphorous 10.1 mg kg⁻¹, available potassium 70.5 mg kg⁻¹, sand (0.02–2 mm) 57.8 %, silt (0.002–0.02 mm) 28.5 %, clay (<0.002 mm) 13.7 % and pH 7.9 (water).

The early-maturing late japonica rice (*Oryza sativa* L.) cultivar Wuxiangjing 14 was planted in this study. The rice seeds were sown in a separate seedling bed in mid-May and transplanted into the experimental plots in mid-June. The seedlings in the ambient plots were grown under the ambient CO₂ concentration, while those in the FACE plots were grown under the elevated CO₂ concentration. The seedlings were manually transplanted at a density of 3 seedlings hill⁻¹ and 24 hills m⁻² and harvested in mid- to late October. Each rice season was followed by a winter wheat (*Triticum aestivum* L.) growing period, which lasted from mid-November until the following late May. The fields were left fallow for approximately 2 weeks between

the two cropping seasons. The farming practices (e.g., water management and the application of pesticides and herbicides) were applied following the local farmers' traditions. The fields were irrigated 2 or 3 days before rice transplanting, and a 1-week long mid-season drainage occurred at approximately 30 days after transplanting (DAT). After the drainage, the fields were intermittently irrigated until the final drainage, which occurred approximately 3–4 weeks before the harvest. No irrigation was adopted after the last drainage, and this lasted until the submergence for the cultivation of rice in the following year.

FACE system

The experiment was organized to facilitate completely randomized replicates with split–split subplots. The atmospheric CO₂ concentration was treated as the main effect. The nitrogen addition was the split-plot treatment, and crop residue was the split–split plot treatment. The rice FACE system has six plots—three plots were randomly allocated for the elevated CO₂ treatments ($200 \pm 40 \mu\text{mol mol}^{-1}$ higher than the ambient level, hereinafter referred to as FACE) and three for the ambient treatments ($375 \mu\text{mol mol}^{-1}$ on average). In the FACE plots, the plants were grown within 12 m diameter 'rings' in which pure CO₂ gas was released from peripheral emission tubes set 0.5 m above the

canopy. A nominal usable area of each plot was ca. 80 m² with a 1 m buffer zone around the emission tubes. The ambient plots had no ring structures, and the plants were grown under ambient CO₂ condition. The FACE treatment was applied for daylight hours in both the rice and wheat growing seasons, and from mid-June 2004 to late October 2007. The FACE treatment was, however, occasionally paused during short fallow periods (approximately 2 weeks) between the two crop-growing seasons. Details of the design, rationale, operation, and performance of the FACE system can be found in previous reports (Liu et al. 2002; Okada et al. 2001). Each CO₂ plot was equally split into two subplots applied with different levels of nitrogen fertilizers. In each nitrogen subplot, three levels of crop residue subplots were randomly assigned.

Nitrogen and crop residue additions

Ammonium-based nitrogen fertilizers (urea plus compound fertilizer—N:P₂O₅:K₂O = 15 %:15 %:15 %) and wheat residues that were harvested in the preceding crop season were applied in the rice seasons. Four levels of nitrogen addition rates (0, 125, 250 and 350 kg N ha⁻¹, which are hereinafter referred to as N0, N125, N250 and N350, respectively) were applied for both the FACE and ambient treatments in the rice season (Table 1); N125 and N250 were applied in 2004

Table 1 Field treatments of the nitrogen fertilizer (kg N ha⁻¹)

| Years | Residue codes | Fertilizer codes ^a | Total fertilizer nitrogen | Basal fertilizer ^b | Tillering fertilizer ^c | Earing fertilizer ^d |
|-------|---------------|-------------------------------|---------------------------|-------------------------------|-----------------------------------|--------------------------------|
| 2004 | HR, MR, ZR | N250 | 250 | 90 | 60 | 100 |
| | | N125 | 125 | 75 | 0 | 50 |
| 2005 | HR, MR, ZR | N250 | 250 | 90 | 60 | 69 |
| | | N125 | 125 | 75 | 0 | 24 |
| 2006 | HR, MR, ZR | N350 | 350 | 126 | 84 | 140 |
| | | N0 | 0 | 0 | 0 | 0 |
| 2007 | ZR | N350 | 350 | 126 | 84 | 140 |
| | | N0 | 0 | 0 | 0 | 0 |

The organic matter additions and their referring codes

HR high residue rate (2,000 kg C ha⁻¹), *MR* moderate residue rate (1,000 kg C ha⁻¹), *ZR* no residue added

^a The codes N0, N125, N250 and N350 indicate the nitrogenous fertilizer application of 0, 125, 250 and 350 kg N ha⁻¹, respectively, during the rice season

^b The basal fertilizer was applied on 15 June (i.e., 1 day prior to transplanting)

^c The tillering fertilizer was applied on 21 June

^d The earing fertilizer was applied on 28 July

and 2005, and N0 and N350 were applied in 2006 and 2007. A replicate nitrogen subplot was approximately 40 m². The time and amount of nitrogen application in various nitrogen subplots can be found in Table 1. Phosphorous and potassium fertilizers were basally applied in all of the subplots at the rates of 33 kg P ha⁻¹ and 62 kg K ha⁻¹ in each rice season. The nitrogen fertilizer types were the same in all the experiment years. For N0 subplots in 2006 and 2007, phosphorous fertilizer was Ca(H₂PO₄)₂ and potassium fertilizer was KCl. For all the other nitrogen subplots, phosphorous and potassium fertilizers were from compound fertilizers (N:P₂O₅:K₂O = 15 %:15 %:15 %).

For each nitrogen subplot, three wheat residue incorporation rates (0, 1,000 and 2,000 kg C ha⁻¹, hereinafter referred to as ZR, MR and HR, respectively) were applied in 2004–2006. In 2007, however, only ZR was applied. The field area of each residue treatment replicate was at least 1 m². The winter wheat stubble, the height of which was approximately 5–10 cm, remained for all of the field plots. One day before submergence, the crop residues (harvested wheat straw from the previous crop season that was cut to 10 cm long) of the MR and HR treatments were incorporated into the cultivation layer of the sub-subplots for all of the nitrogen treatments. In the previous winter wheat seasons, the rice stubble (5–10 cm high) remained in the fields. No additional crop residues or manure from other sources was incorporated in the rice–wheat rotation.

Measurement of the aboveground biomass

At the harvest, the aboveground rice biomass was measured by drying at 80 °C for 48 h.

Measurement of CH₄ flux

The measurements of the CH₄ emissions in situ were performed in the rice seasons of 2004–2007. The methane fluxes from each sub-subplot were measured once every 3–4 days using static, opaque chamber methods in combination with gas chromatography techniques (Wang and Wang 2003; Xu et al. 2004; Zheng et al. 2006). A stainless-steel frame, which covered an area of 0.25 m² and six hills of rice, was permanently installed in the center of each sub-subplot. A 0.5 or 1 m high, gas-tight chamber was temporarily installed on the frame when sampling

occurred. To determine the CH₄ flux, five gas samples were taken with 60-ml plastic syringes at 5 min intervals. The sampling process was completed between 0800 and 1100 LST, at which time the measured flux was expected to accurately represent the daily mean emission (Zheng et al. 1998). Immediately after taking the fifth gas sample, the chamber was removed from the frame. Within at most 10 h of sampling, the gas samples that were stored in the syringes were analyzed using a gas chromatograph with a flame ionization detector (FID; Wang and Wang 2003; Xu et al. 2004) in a temporary laboratory that was located near the experimental site.

Statistical analysis

For analyses of differences of CH₄ daily fluxes and the aboveground rice biomasses on a single year basis, a split-plot design was used, with CO₂ levels as the main plots, nitrogen levels as the subplots, and crop residue as the sub-subplots. One-way ANOVA was used to evaluate the CO₂ effect on rice biomasses. One-way repeated measures ANOVA was applied to test the CO₂ effect when all data of daily CH₄ fluxes were grouped in 2 categories of FACE and ambient. The regulations of nitrogen fertilization and crop residue incorporation on the CO₂ effects on seasonal cumulative CH₄ emissions were determined by paired *t* test. The levels of *p* ≥ 0.10, <0.10, <0.05 and <0.01 were considered to be not significant, slightly significant, significant and highly significant, respectively.

Results

Aboveground rice biomass

In all of the sub-subplots during 2004–2007, with the exception of N0-HR in 2006, the aboveground rice biomass at the time of harvest was enhanced by the elevated CO₂ level (Fig. 1). The mean increase is about 23 %, with a range from 3 to 43 %. Among the totally 20 pairs of treatments, however, only 11 pairs showed significant CO₂-induced increases in aboveground rice biomass (Fig. 1). Both the crop residue and the nitrogen fertilizer incorporation slightly altered the effects of the elevated atmospheric CO₂ on the rice biomass. In the ZR sub-subplots with various nitrogen addition rates, the effect of the

elevated atmospheric CO₂ on the rice biomass was 27 % on average, whereas the CO₂ effects in MR and HR reduced to 14 and 13 %, respectively. In N0, N125, N250 and N350, the stimulations of the elevated atmospheric CO₂ on the rice biomass were 12, 16, 24 and 19 %, respectively.

CH₄ flux

The seasonal dynamic patterns of the CH₄ fluxes were very similar under the FACE and ambient conditions, but they varied in different years (Fig. 2). For the HR and MR sub-subplots, the CH₄ fluxes rapidly increased after transplanting and remained high until the mid-season drainage at approximately 30 DAT. During the following intermittent irrigation periods, CH₄ was emitted at relatively high rate until approximately 90 DAT in 2004, 60 DAT in 2005, and 105 DAT in 2006. After this time, the CH₄ fluxes remained close to zero. For the ZR sub-subplots, the CH₄ fluxes were much lower than those of HR and MR, but the seasonal patterns in 2004–2006 were similar to those of HR and MR. In 2007, however, the CH₄ fluxes from the ZR sub-subplots gradually increased until approximately 60 DAT and then gradually decreased. The average CH₄ fluxes in a single year varied largely with the different rates of the organic matter addition, but showed only slight differences between the two CO₂ or nitrogen levels. The ranges of the CH₄ fluxes were 8–15, 5–12, and

Fig. 2 Methane (CH₄) fluxes that were observed in the plots under elevated (FACE) and ambient (Ambient) CO₂ conditions. The averages of three replicates are provided. The vertical error bars represent standard errors. The definitions of N0, N125, N250, N350, ZR, MR and HR are provided in the footnotes of Table 1 and in the text

2–8 mg CH₄-C m⁻² h⁻¹ for HR, MR and ZR, respectively. The CH₄ fluxes from the MR or ZR sub-subplots showed no significant inter-annual differences in 2005–2007, while the CH₄ fluxes from the HR sub-subplots showed significant inter-annual differences in 2004–2007 rice seasons.

Effects of the elevated CO₂ level on the seasonal cumulative CH₄ emission

Table 2 presents the seasonal cumulative CH₄ emissions from the paddy rice fields under the FACE and ambient conditions. With various nitrogen fertilizer or crop residue incorporation rates, the seasonal CH₄ emissions varied largely; they ranged from 72 to 445 kg CH₄-C ha⁻¹ and 94–499 kg CH₄-C ha⁻¹ for the ambient and elevated CO₂ treatments, respectively. Sixteen of the 20 pairs of CO₂ treatments showed increases of CH₄ emissions that were induced by the elevated CO₂, whereas the other data showed slight decreases. Based upon the average measurements of all the sub-subplots, the elevated atmospheric CO₂ increased the seasonal cumulative CH₄ emissions

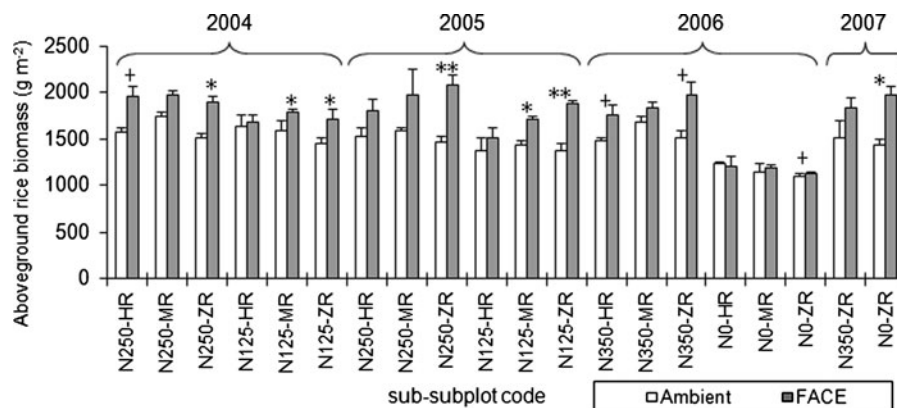


Fig. 1 The aboveground rice biomass (g m⁻²) at the harvest in 2004–2007. The given data are the means of three replicates under the elevated (FACE) or ambient CO₂ (Ambient) conditions. The definitions of N0, N125, N250, N350, ZR, MR and HR are provided in the footnotes of Table 1 and in the

text. The symbols +, *, and ** indicate the significant levels of $p < 0.10$, $p < 0.05$, and $p < 0.01$ respectively, for the differences between the two CO₂ levels, while no symbol indicates no significance ($p \geq 0.10$) in the ANOVA test

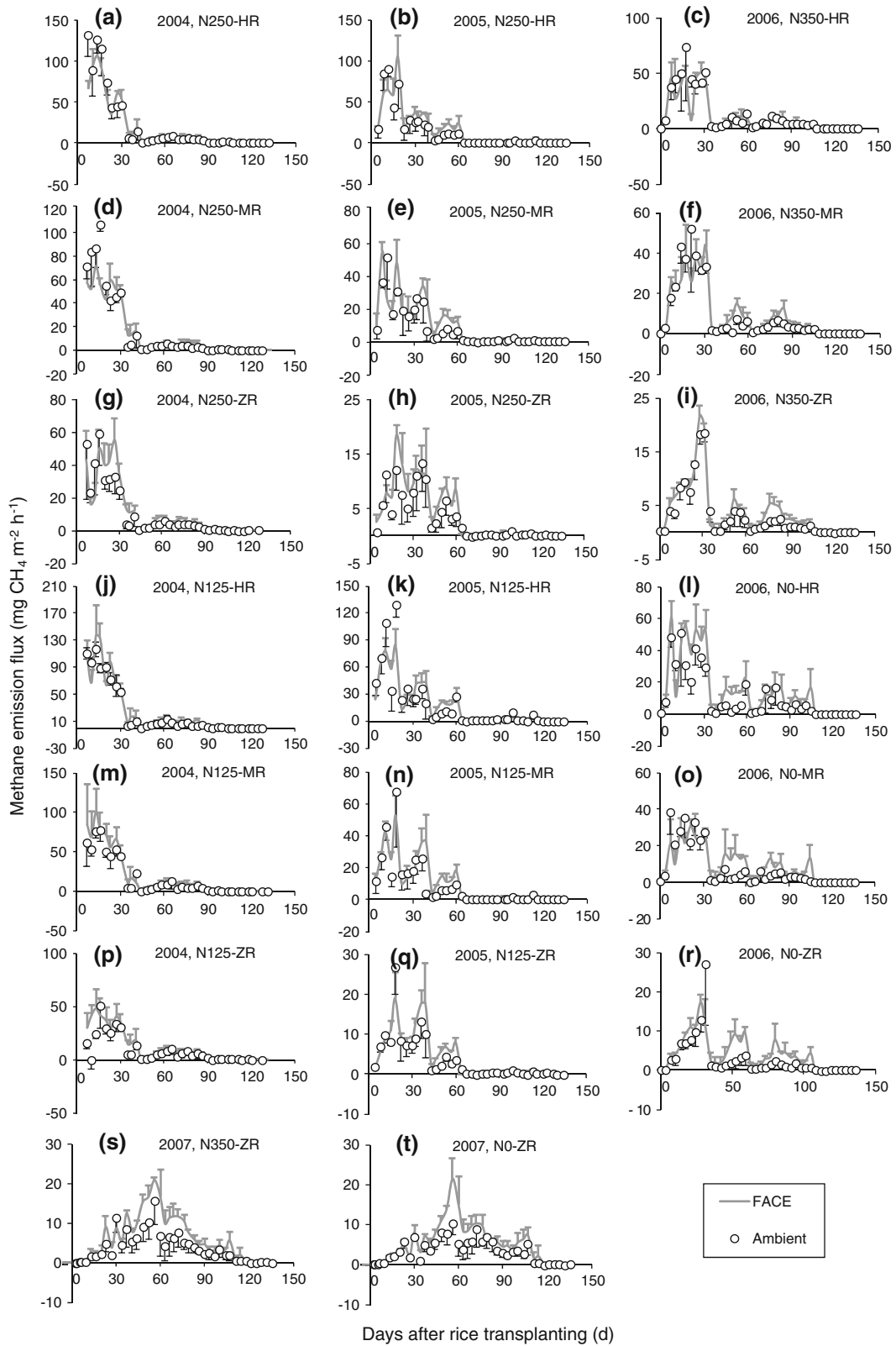


Table 2 Seasonal cumulative methane (CH₄) emissions (SME) from the paddy rice fields that were exposed to the elevated carbon dioxide (F) or the ambient (A) condition and their relative changes that were induced by the elevated carbon dioxide

| Years | Code | SME(kg CH ₄ -C ha ⁻¹) | | |
|-------|---------|--|----------|-------------|
| | | F | A | Changes (%) |
| 2004 | N250-HR | 392 (40) | 425 (45) | -8 (16) |
| | N250-MR | 319 (22) | 347 (42) | -8 (12) |
| | N250-ZR | 243 (36) | 205 (52) | 18 (20) |
| | N125-HR | 499 (113) | 445 (26) | 12 (34) |
| | N125-MR | 410 (120) | 325 (56) | 26 (51) |
| | N125-ZR | 261 (47) | 180 (51) | 45 (69) |
| 2005 | N250-HR | 379 (41) | 316 (76) | 20 (16) |
| | N250-MR | 233 (38) | 182 (61) | 28 (22) |
| | N250-ZR | 94 (17) | 72 (24) | 30 (42) |
| | N125-HR | 388 (49) | 396 (56) | -2 (11) |
| | N125-MR | 237 (30) | 193 (45) | 23 (29) |
| | N125-ZR | 98 (11) | 79 (16) | 24 (35) |
| 2006 | N350-HR | 268 (45) | 317 (84) | -16 (38) |
| | N350-MR | 229 (55) | 219 (32) | 4 (31) |
| | N350-ZR | 92 (5) | 74 (8) | 24 (17) |
| | N0-HR | 369 (67) | 259 (53) | 42 (63) |
| | N0-MR | 222 (47) | 184 (18) | 21 (29) |
| | N0-ZR | 100 (13) | 66 (17) | 52 (87) |
| 2007 | N350-ZR | 149 (26) | 93 (37) | 61 (37) |
| | N0-ZR | 133 (25) | 85 (23) | 57 (28) |

The definitions of N0, N125, N250, N350, ZR, MR and HR are provided in the footnotes of Table 1 and in the text. The values that are outside of the parentheses are the means of three replicates, and those inside the parentheses are the standard errors. The relative changes were calculated by $(F - A)/A \times 100$

by 15 %. However, the difference was not statistically significant. A significant interaction between N fertilization and elevated CO₂ was found in 2007, however, there were no other significant interactions among the atmospheric CO₂, N and crop residue incorporations (Table 3).

Regulation of the crop residue and nitrogen fertilizer addition on the effect of elevated CO₂

The data in Table 2 clearly indicate that the effect of the elevated CO₂ on the seasonal cumulative CH₄ emissions was affected by crop residue incorporation. This regulation is further interpreted in Fig. 3a. The CO₂ effect became weaker with the increasing residue addition rates. On average, the CO₂ effects were 37 ($p < 0.01$), 14 ($p < 0.1$) and 6 % ($p > 0.1$) for ZR, MR and HR, respectively. The significance level of the

CO₂ effect also decreased with the increasing residue addition rates (Fig. 3a).

As shown in Fig. 3b, nitrogen fertilizer seemed to inhibit the effect of elevated CO₂ on the seasonal cumulative CH₄ emissions. The highest stimulation of the elevated CO₂ on CH₄ emissions occurred in the N0 subplots, in which the CO₂ effect was 37 % ($p < 0.05$). In the N125, N250 and N350 subplots, the averaged CO₂ effects reduced to 17 ($p < 0.05$), 7 ($p > 0.1$) and 5 % ($p > 0.1$), respectively.

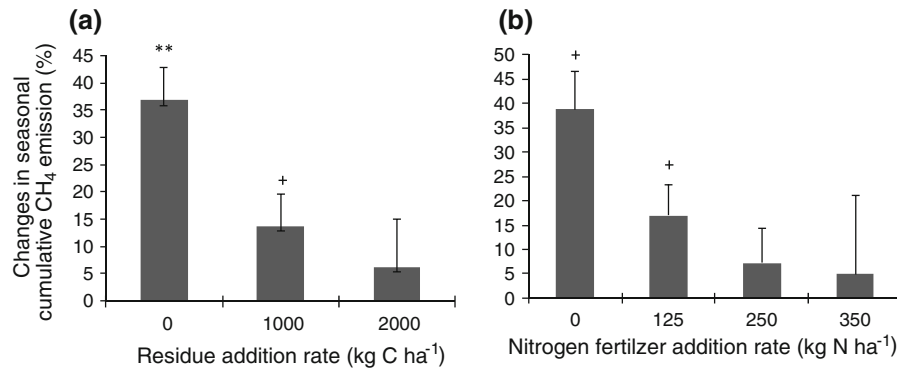
Discussion

High amount of CH₄ emission in 2004 rice growing season

The addition of organic matters such as crop residue is one of the major methanogenic substrates in rice

Table 3 The significance levels (*p* values) of CO₂, nitrogen (N), residue (R), and their interactions on CH₄ emission from 2004 to 2007

| Years | CO ₂ | N | R | CO ₂ *N | CO ₂ *R | N*R | CO ₂ *N*R |
|-------|-----------------|-------|-------|--------------------|--------------------|-------|----------------------|
| 2004 | 0.643 | 0.636 | 0.004 | 0.392 | 0.924 | 0.771 | 0.967 |
| 2005 | 0.389 | 0.581 | 0.000 | 0.722 | 0.955 | 0.879 | 0.913 |
| 2006 | 0.341 | 0.992 | 0.000 | 0.227 | 0.997 | 0.825 | 0.514 |
| 2007 | 0.170 | 0.000 | | 0.016 | | | |

**Fig. 3** The relative changes in the seasonal cumulative CH₄ emission that were caused by the elevated CO₂: **a** the crop residue treatment, **b** the nitrogen treatment. The error bars represent the standard errors. The symbols + and ** indicate

the significant levels of $p < 0.10$ and $p < 0.01$, respectively, while *no symbol* indicates no significance ($p \geq 0.10$) for the differences between the two CO₂ levels

paddies (Huang et al. 2004; Zheng et al. 2006). Thus, we checked the inter-annual differences of CH₄ emission with the same residue addition rate. There were no significant inter-annual differences in CH₄ emissions among 2005 to 2007. However, the seasonal cumulative CH₄ emissions in 2004 were larger than the means of the following 3 years by 31, 65 and 136 % for HR, MR, and ZR, respectively. There was a special farm management in 2004 different from 2005–2007. The straws of previous winter wheat were removed from the fields before rice transplanting in 2005–2007. However, in 2004, i.e. the first rice season of the FACE study, the straws of previous winter wheat were burned in the field and the ashes were ploughed into soil 2 days before submergence. The in situ combustion of straws may contribute to the high rate of CH₄ emission in 2004. Ma et al. (2008) found a similar result that in situ combustion of straws could enhance CH₄ emission from paddy rice fields by 130 %. They suggested that the ashes from the wheat straws contained much organic carbon which

could be utilized by methanogens, resulting in increased CH₄ emission.

Effects of the elevated CO₂ on CH₄ emission from intermittently irrigated rice fields

The emission of CH₄ from irrigated rice fields is controlled by the production of CH₄ as well as oxidation and transport processes, all of which can be affected by elevated atmospheric CO₂. Most of the published environment-controlled or FACE studies have consistently demonstrated that elevated atmospheric CO₂ could significantly stimulate CH₄ emissions from paddy rice fields (Cheng et al. 2006; Inubushi et al. 2003; Xu et al. 2004; Zheng et al. 2006; Ziska et al. 1998). Analyzing both previous studies and their own studies, Zheng et al. (2006) suggested that elevated CO₂ affected CH₄ emission from paddy rice fields primarily by stimulating rice growth.

The 15 % stimulation that was observed in the present study is much weaker than the previous FACE

studies (Inubushi et al. 2003; Tokida et al. 2010; Zheng et al. 2006). A previous FACE study that was also conducted in the Yangtze River Delta showed significant stimulation of CH₄ emissions from 53 to 188 % (Zheng et al. 2006). A major difference is that the previous study was conducted in clay paddies, whereas the present study was conducted in sandy loam paddies. The hydraulic conductivity of sandy loam is usually higher than that of clay soil, which requires more irrigation to meet the rice growth demand. In fact, the field in the present study was intermittently irrigated, which might result in higher soil redox potential (Eh) than long-term flooded soil (private communication with Dr. Xiaozhi Wang of Yangzhou University). Soil Eh is one of the most sensitive environmental factors that control CH₄ production. The relatively high soil Eh can remarkably inhibit CH₄ production (Mitra et al. 2002a, b; Wang 2001). Therefore, the different soil type may alter the effect of the elevated CO₂ on the CH₄ emission.

When we statistically tested the effect of elevated CO₂ for each pair of the elevated and ambient CO₂ treatments, it shows the stimulation is not significant. The reasons may include the following aspects.

Firstly, as mentioned above, the elevated CO₂ indirectly stimulates CH₄ emission primarily via intensifying rice growth. In the present study, probably due to the uncontrolled rice diseases (ex. rice stripe Tenuivirus) and inconsistent irrigation frequencies and water levels, nearly half of the total 20 pairs of sub-subplots showed no significant differences in the aboveground rice biomass between the two CO₂ levels (Fig. 1). Consequently, the relevant factors or processes (methanogenic substrates, methanogen and methanotroph activity, vascular CH₄ transport) controlled by rice growth may not significantly vary between the two CO₂ levels, which resulted in small and insignificant differences in CH₄ emission from the FACE and ambient fields.

Methane is produced by methanogens under anaerobic conditions, and is oxidized to CO₂ or assimilated into the microbial biomass by methanotrophs under aerobic conditions. Therefore, the populations of methanogens and methanotrophs are key regulators in CH₄ emission (Conrad 1999; Kruger and Frenzel 2003; Kruger et al. 2001; Yao and Conrad 1999). In the FACE study that was conducted in Wuxi in the Yangtze River Delta (Zheng et al. 2006), the methanogen populations were significantly increased by

197 ± 71 %, whereas the methanotrophic populations only changed by 2 ± 16 % (Yue et al. 2007), which contributed largely to the stimulation of CH₄ emission by elevated CO₂ (Zheng et al. 2006). Using the most probable number (MPN) method as Yue et al. (2007), Wang et al. (2006) measured the populations of methanogens and methanotrophs in the fresh soils of four sub-subplots (2 HR and 2 ZR) in the present experimental fields in 2005 rice season. They found that the elevated CO₂ had no significant impacts on populations of both methanogens and methanotrophs, which may be the major microbial reason causing a weaker stimulation of CH₄ emissions by elevated CO₂.

Similar to our results, a recent FACE study in Japan found that the enhancement (+26 %) in CH₄ emission resulting from the elevated atmospheric CO₂ (ambient + 200 μmol mol⁻¹) was statistically insignificant (Tokida et al. 2010). They speculated that the absence of a significant stimulatory effect of FACE on CH₄ emission could be attributable partly to the heterogeneity of the soil within the experimental sites. Obviously, more studies are needed to understand the underlying mechanisms responsible for the large variation in the effects of elevated CO₂ on CH₄ emissions from study to study.

The statistical power of ANOVA in this study was very low. Three replications of FACE rings only gave a degree of freedom for all effect of 1 or 2 for the error. Therefore, there was a high risk of type II error—falsely rejecting the null hypothesis (no difference between FACE and ambient), i.e., in our study an actual simulating effect of elevated CO₂ on CH₄ emission from rice paddies was probably disregarded by the test. Although the statistical test failed to reject the null hypothesis, but never proved that there was no difference between FACE and ambient treatments. In general, increasing the sample size can reduce type II error. We collectively tested the effect of elevated CO₂ by splitting all data into two categories, FACE and ambient, as treating the sub treatments as repeated measurements in different conditions. This analysis increased the effect size to 120 (20 pairs F-A with 3 repetitions) and consequently indicated that elevated CO₂ significantly stimulated CH₄ emission. However, in experimented fields, the effort to increase sample size generally may not be feasible. Therefore, many of the effect, even though having a rather high effect size, were not significant because of type II error, like our split-plot ANOVA test.

Impacts of the crop residue and nitrogen fertilizer incorporation on CH₄ emission in response to elevated CO₂

To our knowledge, there is only one published report that focuses on the impacts of crop residue and nitrogen fertilizer incorporation on CH₄ emissions in response to elevated CO₂ (Zheng et al. 2006). The present study showed that crop residue incorporation appeared to inhibit the stimulatory effect of elevated CO₂ on CH₄ emission. An important reason for this result may be related to the different elevated CO₂-induced increases of rice biomass in various crop residue treatments. In the ZR sub-subplot, the elevated CO₂ stimulated the aboveground rice biomass by 26 %, which could provide a large increase in the methanogenic substrates. However, the increments of the rice biomass induced by the elevated CO₂ dropped to 12 % in the MR and HR sub-subplots, where the increased proportions of the methanogenic substrates would be lower than in ZR, thus resulting in weaker stimulations of CH₄ emissions by elevated CO₂.

Zheng et al. (2006) also reported that the significant stimulation of CH₄ emissions by the elevated CO₂ correlated negatively with the addition rates of the organic matter. They suggested that this result might be due to an increased C/N ratio in the incorporated wheat straw that was produced under the elevated CO₂. The increased C/N ratio would limit the decomposition of the residue, and it would therefore result in a decrease of the methanogenic substrates. Liu et al. (2009) reported an incubation study that focused on the decomposition process of crop residue that was mixed with a paddy soil. The authors used winter wheat and rice straw that was produced under ambient and elevated CO₂ conditions in the same FACE station as reported by Zheng et al. (2006). They found that there were no significant differences in the straw-induced CH₄ emissions of the incorporated wheat straw that were produced either under ambient or under elevated CO₂ conditions. It should be noted that the soil in the incubation was not taken from the same FACE station but rather from an experimental station of the International Rice Research Institute; furthermore, the incubation experiment was not performed under elevated CO₂ conditions. Consequently, the incubation results may vary largely from the in situ FACE experiment. Therefore, appropriate experiments are necessary to

understand the elevated atmospheric CO₂ influence on the decomposition of organic matter that is incorporated into soil.

Zheng et al. (2006) reported that the significant stimulation of CH₄ emissions by the elevated CO₂ correlated positively with nitrogen addition rates. They suggested that low soil nitrogen availability can counteract the stimulatory effect of elevated CO₂ on CH₄ emissions from nitrogen-poor paddy rice ecosystems, whereas high soil nitrogen availability can further amplify the stimulatory effect of elevated CO₂ on CH₄ emission from nitrogen-rich paddy rice ecosystems. In the present study, the elevated CO₂ significantly stimulated CH₄ emissions from the fields that lacked nitrogen fertilizer, and the addition of nitrogen fertilizer seemed to hinder the stimulatory effects of the elevated CO₂ on the CH₄ emission. However, the significant regulation of the nitrogen incorporation occurred only in the N125 treatment. In different nitrogen treatments, there were no significant differences in the elevated CO₂ effects on the aboveground rice biomass, which probably resulted in the similar elevated CO₂-induced methanogenic substrates and vascular transport capacities. Consequently, the CO₂-induced CH₄ emissions in the various nitrogen treatments were similar. Wang et al. (2006) reported that there were no significant differences in the CO₂-induced methanogen or methanotroph population changes in the N125 and N250 treatments, which may also support the insignificant differences in the elevated CO₂-induced CH₄ emission between the two nitrogen levels.

Conclusion

Our results revealed that an elevated atmospheric CO₂ concentration ($200 \pm 40 \mu\text{mol mol}^{-1}$ higher than the ambient atmosphere) stimulated the seasonal cumulative CH₄ emissions by an average of 15 % from the paddy rice fields with a sandy loam texture. The stimulatory extent was smaller than those of previous studies, which might result from different soil types or farm management practices. In addition, the present study also implied that the stimulatory effect of the elevated CO₂ on CH₄ emission tended to be inhibited by the incorporation of nitrogen fertilizer and crop residue. To accurately evaluate the effects of increasing atmospheric CO₂ concentration on CH₄ emissions

from wetland ecosystems, other factors such as soil types and farm managements must be considered.

Acknowledgments We are grateful to the financial supports from the Ministry of Science and Technology of China (grant number: 2012CB417100), the National Natural Science Foundation of China (grant numbers: 41021004, 40425010) and the Chinese Academy of Sciences (grant number: KZCX3-SW-440), and to the technical supports from NIAES and NARO, Ministry of Agriculture, Forestry and Fisheries, Japan. We also thank Y. Han, L.-X. Yang, Y.-S. Wang, G. Liu, C.-Y. Liu, J.-X. Gu, R. Wang, J. Deng and H.-J. Tong for their contributions in the field measurements, technical assistances and fruitful help in preparing for this paper.

References

- Allen LH, Albrecht SL, Colon-Guasp W, Covell SA, Baker JT, Pan DY, Boote KJ (2003) Methane emissions of rice increased by elevated carbon dioxide and temperature. *J Environ Qual* 32:1978–1991
- Bodelier PLE, Laanbroek HJ (2004) Nitrogen as a regulatory factor of methane oxidation in soils and sediments. *FEMS Microbiol Ecol* 47:265–277. doi:10.1016/s0168-6496(03)00304-0
- Cai Z, Shan Y, Xu H (2007) Effects of nitrogen fertilization on CH₄ emissions from rice fields. *Soil Sci Plant Nutr* 53: 353–361
- Cassman KG, Peng S, Oik DC, Ladha JK, Reichardt W, Dobermann A, Singh U (1998) Opportunities for increased nitrogen-use efficiency from improved resource management in irrigated rice systems. *Field Crops Res* 56:7–39
- Cheng WG, Inubushi K, Yagi K, Sakai H, Kobayashi K (2001) Effects of elevated carbon dioxide concentration on biological nitrogen fixation, nitrogen mineralization and carbon decomposition in submerged rice soil. *Biol Fertil Soils* 34:7–13
- Cheng WG, Yagi K, Sakai H, Kobayashi K (2006) Effects of elevated atmospheric CO₂ concentrations on CH₄ and N₂O emission from rice soil: an experiment in controlled-environment chambers. *Biogeochemistry* 77:351–373. doi:10.1007/s10533-005-1534-2
- Conrad R (1999) Contribution of hydrogen to methane production and control of hydrogen concentrations in methanogenic soils and sediments. *FEMS Microbiol Ecol* 28: 193–202
- Conrad R (2002) Control of microbial methane production in wetland rice fields. *Nutr Cycl Agroecosyst* 64:59–69
- Dacey JWH, Drake BG, Klug MJ (1994) Stimulation of methane emission by carbon dioxide enrichment of marsh vegetation. *Nature* 370:47–49
- Denier van der Gon HAC, Neue HU (1995) Influence of organic matter incorporation on the methane emission from a wetland rice field. *Global Biogeochem Cycles* 9:11–22
- Hoque MM, Inubushi K, Miura S, Kobayashi K, Kim HY, Okada M, Yabashi S (2001) Biological dinitrogen fixation and soil microbial biomass carbon as influenced by free-air carbon dioxide enrichment (FACE) at three levels of nitrogen fertilization in a paddy field. *Biol Fertil Soils* 34:453–459
- Houghton JT, Ding Y, Griggs D, Noguer M, Van der Linden P, Dai X, Maskell K, Johnson C (2001) The scientific basis; contribution of working group I to the third assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge
- Huang Y, Zhang W, Zheng XH, Li J, Yu YQ (2004) Modeling methane emission from rice paddies with various agricultural practices. *J Geophys Res Atmos* 109. doi:10.1029/2003jd004401
- Hutchin PR, Press MC, Lee JA, Ashenden TW (1995) Elevated concentrations of CO₂ may double methane emissions from mires. *Glob Change Biol* 1:125–128
- Inubushi K, Cheng WG, Aonuma S, Hoque MM, Kobayashi K, Miura S, Kim HY, Okada M (2003) Effects of free-air CO₂ enrichment (FACE) on CH₄ emission from a rice paddy field. *Glob Change Biol* 9:1458–1464
- IPCC (2001) Climate change 2001: The scientific basis. In: Houghton RA et al. (eds) Contribution of working group I to the third assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge
- IPCC (2007) Climate change 2007: the physical science basis. Cambridge University Press, Cambridge
- Kim HY, Lieffering M, Miura S, Kobayashi K, Okada M (2001) Growth and nitrogen uptake of CO₂-enriched rice under field conditions. *New Phytol* 150:223–229
- Kim HY, Lieffering M, Kobayashi K, Okada M, Mitchell MW, Gumpertz M (2003) Effects of free-air CO₂ enrichment and nitrogen supply on the yield of temperate paddy rice crops. *Field Crops Res* 83:261–270
- Kimball BA (1992) Cost comparisons among free-air CO₂ enrichment, open-top chamber, and sunlit controlled environment chamber methods of CO₂ exposure. *Crit Rev Plant Sci* 11:265–270
- Kludze HK, Delaune RD, Patrick WH (1993) Aerenchyma formation and methane and oxygen exchange in rice. *Soil Sci Soc Am J* 57:386–391
- Kruger M, Frenzel P (2003) Effects of N-fertilisation on CH₄ oxidation and production, and consequences for CH₄ emissions from microcosms and rice fields. *Glob Change Biol* 9:773–784
- Kruger M, Frenzel P, Conrad R (2001) Microbial processes influencing methane emission from rice fields. *Glob Change Biol* 7:49–63
- Le Mer J, Roger P (2001) Production, oxidation, emission and consumption of methane by soils: a review. *Eur J Soil Biol* 37:25–50. doi:10.1016/s1164-5563(01)01067-6
- Liu G, Han Y, Zhu J, Okada M, Nakamura H, Yoshimoto M (2002) Rice-wheat rotational FACE platform. I. System structure and control. *Chin J Appl Ecol* 13:1253–1258 (in Chinese with English abstract)
- Liu J, Han Y, Cai ZC (2009) Decomposition and products of wheat and rice straw from a FACE experiment under flooded conditions. *Pedosphere* 19:389–397
- Long SP, Ainsworth EA, Leakey ADB, Nosberger J, Ort DR (2006) Food for thought: lower-than-expected crop yield stimulation with rising CO₂ concentrations. *Science* 312:1918–1921. doi:10.1126/science.1114722

- Ma J, Xu H, Yagi K, Cai ZC (2008) Methane emission from paddy soils as affected by wheat straw returning mode. *Plant Soil* 313:167–174. doi:[10.1007/s11104-008-9689-y](https://doi.org/10.1007/s11104-008-9689-y)
- Mitra S, Wassmann R, Jain MC, Pathak H (2002a) Properties of rice soils affecting methane production potentials: 1. Temporal patterns and diagnostic procedures. *Nutr Cycl Agroecosyst* 64:169–182
- Mitra S, Wassmann R, Jain MC, Pathak H (2002b) Properties of rice soils affecting methane production potentials: 2. Differences in topsoil and subsoil. *Nutr Cycl Agroecosyst* 64:183–191. doi:[10.1023/a:1021175404418](https://doi.org/10.1023/a:1021175404418)
- Okada M, Lieffering M, Nakamura H, Yoshimoto M, Kim HY, Kobayashi K (2001) Free-air CO₂ enrichment (FACE) using pure CO₂ injection: system description. *New Phytol* 150:251–260
- Schimel J (2000) Global change—rice, microbes and methane. *Nature* 403:375–377
- Silvola J, Saarnio S, Foot J, Sundh I, Greenup A, Heijmans M, Ekberg A, Mitchell E, van Breemen N (2003) Effects of elevated CO₂ and N deposition on CH₄ emissions from European mires. *Global Biogeochem Cycles* 17:1068. doi:[10.1029/2002gb001886](https://doi.org/10.1029/2002gb001886)
- Tokida T, Fumoto T, Cheng W, Matsunami T, Adachi M, Katayanagi N, Matsushima M, Okawara Y, Nakamura H, Okada M, Sameshima R, Hasegawa T (2010) Effects of free-air CO₂ enrichment (FACE) and soil warming on CH₄ emission from a rice paddy field: impact assessment and stoichiometric evaluation. *Biogeosciences* 7:2639–2653. doi:[10.5194/bg-7-2639-2010](https://doi.org/10.5194/bg-7-2639-2010)
- Wang M (2001) Methane emission from Chinese rice paddies. Science Press, Beijing
- Wang YS, Wang YH (2003) Quick measurement of CH₄, CO₂ and N₂O emissions from a short-plant ecosystem. *Adv Atmos Sci* 20:842–844
- Wang B, Xu Y, Wang Z, Li Z, Ding Y, Guo Y (1999) Methane production potentials of twenty-eight rice soils in China. *Biol Fertil Soils* 29:74–80
- Wang SY, Lin H, Yi S, Bao-ling M, Jian-guo Z (2006) Effects of FACE on population and activities of methanogen and methanotroph in paddy soil. *Soils* 38:768–773 (in Chinese with English abstract)
- Xu ZJ, Zheng XH, Wang YS, Han SH, Huang Y, Zhu JG, Butterbach-Bahl K (2004) Effects of elevated CO₂ and N fertilization on CH₄ emissions from paddy rice fields. *Global Biogeochem Cycles* 18. doi:[10.1029/2004gb002233](https://doi.org/10.1029/2004gb002233)
- Yagi K, Li Z, Xu H (2002) Effect of elevated CO₂ on methane emission from rice fields. In: Proceedings for the 10th CACGP/seventh IGAC conference, September 18–25, Grete
- Yang L, Wang Y, Dong G, Gu H, Huang J, Zhu J, Yang H, Liu G, Han Y (2007) The impact of free-air CO₂ enrichment (FACE) and nitrogen supply on grain quality of rice. *Field Crops Res* 102:128–140
- Yao H, Conrad R (1999) Thermodynamics of methane production in different rice paddy soils from China, the Philippines and Italy. *Soil Biol Biochem* 31:463–473
- Yue J, Shi Y, Zheng XH, Huang GH, Zhu JG (2007) The influence of free-air CO₂ enrichment on microorganisms of a paddy soil in the rice-growing season. *Appl Soil Ecol* 35:154–162. doi:[10.1016/j.apsoil.2006.05.005](https://doi.org/10.1016/j.apsoil.2006.05.005)
- Zheng X, Mingxing W, Yuesi W, Renxing S, Jing L, Heyer J, Kogge M, Laotu L, Jisheng J (1998) Comparison of manual and automatic methods for measurement of methane emission from rice paddy fields. *Adv Atmos Sci* 15:569–579
- Zheng X, Wang M, Wang Y, Shen R, Gou J, Li J, Jin J, Li L (2000) Impacts of soil moisture on nitrous oxide emission from croplands: a case study on the rice-based agro-ecosystem in Southeast China. *Chemos Global Change Sci* 2:207–224
- Zheng X, Zhou Z, Wang Y, Zhu J, Yue J, Shi Y, Kobayashi K, Inubushi K, Huang Y (2006) Nitrogen-regulated effects of free-air CO₂ enrichment on methane emissions from paddy rice fields. *Glob Change Biol* 12:1717–1732
- Ziska LH, Moya TB, Wassmann R, Namuco OS, Lantin RS, Aduna JB, Abao E, Bronson KF, Neue HU, Olszyk D (1998) Long-term growth at elevated carbon dioxide stimulates methane emission in tropical paddy rice. *Glob Change Biol* 4:657–665